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Tuning the incorporation of electroactive metals into titanium phosphate nanoparticles and the reverse metal extraction process: Application as electrochemical labels in multiplex biosensing

Javier Carrasco-Rodríguez <sup>a</sup>, Francisco J. García Alonso <sup>a</sup>, Agustín Costa-García <sup>b</sup>, Daniel Martín-Yerga <sup>b</sup>  

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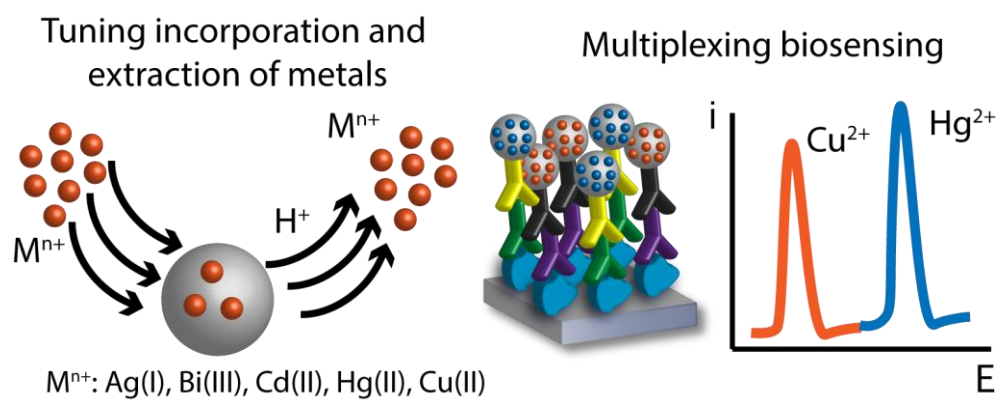
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## GRAPHICAL ABSTRACT



## **HIGHLIGHTS**

- Titanium phosphate nanoparticles were modified with electroactive metals
- Metal amount introduced into the nanoparticles could be tuned
- Acidic media was successful to extract and detect the metals
- Cu and Hg were the most appropriate metals (sensitivity and selectivity)
- A multiplexing biosensor was developed with the modified nanoparticles

1           **Tuning the incorporation of electroactive metals into titanium**  
2           **phosphate nanoparticles and their extraction for multiplexing**  
3                           **electrochemical biosensing**

4

5           *Javier Carrasco-Rodríguez<sup>1</sup>, Francisco J. García Alonso<sup>1</sup>, Agustín Costa-García<sup>2</sup>, Daniel Martín-Yerga<sup>2\*</sup>*

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8           <sup>1</sup>Departamento de Química Orgánica e Inorgánica, Universidad de Oviedo, 33006 Oviedo, Spain.

9           <sup>2</sup>Departamento de Química Física y Analítica, Universidad de Oviedo, 33006 Oviedo, Spain.

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18           \* Corresponding author: Dr. Daniel Martín-Yerga

19           Departamento de Química Física y Analítica

20           Universidad de Oviedo

21           Julián Clavería 8, Oviedo 33006 (Spain)

22           E-mails: dyerga@gmail.com

23           Telephone: (+34) 985103486

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27 **ABSTRACT**

28 In this work, titanium phosphate nanoparticles were modified with different electroactive metals  
29 such as cadmium, bismuth, copper, silver and mercury by a cation exchange reaction. The amount  
30 of metal introduced into the nanoparticles depended strongly on the counter-ion used during the  
31 exchange reaction and the type of metal, so that nanoparticles with a high metallic load could be  
32 generated. For the detection of these metal-modified nanoparticles, the electrolytic medium used  
33 played an important role since the use of acid allows to extract a large part of the introduced metal  
34 by reverting the cation exchange reaction. The electrochemical detection of the nanoparticles was  
35 evaluated, being the nanoparticles modified with copper and mercury the most adequate in terms of  
36 sensitivity and selectivity with the aim of multiplexing detection. As a proof-of-concept, these  
37 nanoparticles were used as a detection label in a multiplexing electrochemical biosensor for the  
38 simultaneous detection of two analytes.

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43 **KEYWORDS:** Nanoparticle; Titanium phosphate; Metal incorporation; Multiplexing; Biosensing

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## 53 INTRODUCTION

54 Metal-based nanoparticles are being widely used as detection label in electrochemical biosensors[1–  
55 3] since they have interesting properties such as the possibility of biofunctionalization, multiple and  
56 easy synthesis methods, biocompatibility and, typically, electroactivity. The great variety of  
57 metallic nanoparticles available is very convenient for multiplexing assays[4,5], where the  
58 simultaneous detection of several analytes is carried out. Nanoparticles that can be loaded with  
59 different electroactive species[6], such as liposomes[7], apoferritins[8,9] or other  
60 nanoparticles[10,11] are also a constant resource in this field. Nevertheless, the development of  
61 novel or enhanced nanoparticles that can load a large amount of electroactive species is still a  
62 relevant research problem in order to improve the sensitivity of the detection in electrochemical  
63 biosensing. For instance, titanium phosphate nanoparticles (TiPNPs)[12,13] are very interesting  
64 because they have a porous structure and an amorphous coating of acid phosphates that provides  
65 them with a strong ionic-exchange functionality. However, these nanoparticles have only been  
66 modified with cadmium, lead or zinc for use as detection labels[14–17]. These metals are reduced at  
67 a quite negative potential, which can also reduce other concomitant species in the solution.  
68 Furthermore, the hydrogen evolution reaction usually occurs at a close potential (in acidic  
69 solutions) and could affect the efficiency of the metal electrodeposition on the electrode surface.  
70 Therefore, the evaluation and characterization of other electroactive metals introduced into these  
71 nanoparticles[18,19] may lead to better detection labels for electrochemical biosensors. In this  
72 work, titanium phosphate nanoparticles were modified with several electroactive metals such as  
73 cadmium, bismuth, silver, mercury and copper. The amount of metal introduced into the  
74 nanoparticles was tuned by changing the metal or the counter-ion used during the cation exchange  
75 reaction. The modification of the nanoparticles with metals that are reduced at more positive metals,  
76 allowed to use different electrolyte media able to extract the metals more efficiently than in  
77 previous studies reported in the literature. The capacity of multiplexing detection of these  
78 nanoparticles was evaluated by the development of a biosensor as a proof-of-concept.

79

## 80 **MATERIALS AND METHODS**

### 81 **Apparatus and electrodes**

82 Electrochemical measurements were conducted with  $\mu$ Stat 8000 (DropSens)  
83 potentiostat/galvanostat interfaced to an Apple Macbook Air laptop and controlled by the  
84 DropView 8400 2.2 software. 8-channel screen-printed carbon electrochemical arrays (SPCEs)  
85 were purchased from DropSens (ref. 8X110). These devices, with a circular working electrode of  
86 2.56 mm diameter, have been previously described[20]. 8-channel arrays were connected to the  
87 potentiostat through a specific connector, DRP-CAST8x. All measurements were carried out at  
88 room temperature and using an aliquot of 25  $\mu$ L of the appropriate solution. All reported potentials  
89 are related to the silver pseudoreference screen-printed electrode.

90

### 91 **Reagents and solutions**

92 Cadmium nitrate, cadmium acetate, cadmium acetylacetonate, mercury acetate, copper acetate,  
93 silver nitrate, bismuth nitrate, bovine serum albumin fraction V (BSA), phosphoric acid(crystalline),  
94 docusate sodium salt(AOT), poly-(allylamine hydrochloride), glutaraldehyde, titanium(IV)  
95 butoxide were purchased from Sigma. Sulfuric acid (98%), acetic acid (100%), phosphoric acid  
96 solution (85%), dried ethanol, sodium hydroxide and hydrochloric acid were purchased from  
97 Merck. Neutravidin (NTV) was purchased from Fisher Scientific. Human tissue transglutaminase  
98 was purchased from Zedira. biotinylated goat anti-human IgA and IgG (anti-IgA-BT, anti-IgG-BT)  
99 were purchased from Life Technologies. Varelista Celikey tissue transglutaminase IgA ELISA kit  
100 was purchased from Phadia. Ultrapure water obtained with a Millipore Direct Q5™ purification  
101 system from Millipore was used throughout this work. All other reagents were of analytical grade.  
102 Unless stated otherwise, 2  $\mu$ L of a nanoparticle dispersion were employed for the modification of  
103 electrodes.

104

## 105 **Synthesis of titanium phosphate nanoparticles modified with metals**

106 The synthesis of titanium phosphate nanoparticles was carried out following a procedure found in  
107 the literature[12] slightly modified and previously reported[18]. For the synthesis of metal-modified  
108 titanium phosphate nanoparticles, an aqueous suspension (1 mL) of TiPNPs (40 mg/mL) was  
109 dispersed in 17 mL of a 10 mM aqueous solution of the appropriate metallic salt and the resulting  
110 mixture was stirred at 50 °C for 24 h. Then, the final mixture was centrifuged, the solid precipitate  
111 was washed three times with 10 mL of ultrapure water and the nanoparticles were dried under  
112 vacuum overnight.

## 113 **Bio-functionalization of MTiPNPs with neutravidin and antibodies**

114 For the bio-functionalization of MTiPNPs, a method previously reported was employed[21]. The  
115 neutravidin-biotin reaction was used for the conjugation of MTiPNPs-NTV with biotinylated anti-  
116 IgA and anti-IgG antibodies. Briefly, in a low-binding micro-tube, a 1:1 mixture (in PBS) of  
117 biotinylated antibody solution (5 µg/mL) and MTiPNPs-NTV (100 µg/mL) were left to incubate for  
118 50 min under constant stirring. After the reaction, a small amount of BSA was added to these  
119 solutions (final concentration of 0.25% BSA), in order to minimize the possible non-specific  
120 adsorptions in the immunosensor.

## 121 **Bioassay and electrochemical detection procedures**

122 For the biosensor, modification of electrodes with the sensing element (transglutaminase) and the  
123 different steps were carried out by following a method previously developed in our group[22] but  
124 using PBS as buffer solution. The reaction with the secondary antibodies was performed using bio-  
125 functionalized MTiPNPs. The electrochemical detection was carried out with 25 µL of 0.1 M  
126 H<sub>2</sub>SO<sub>4</sub> by square-wave anodic stripping voltammetry with a deposition step at -1 V for 60 s.

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## 130 RESULTS AND DISCUSSION

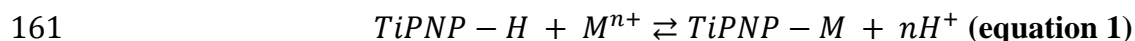
### 131 Tuning the incorporation of metals into titanium phosphate nanoparticles

132 Titanium phosphate nanoparticles modified with metals (MTiPNPs) can be detected by  
133 electrochemical methods after the introduction of electroactive metals into their structure. TiPNPs  
134 modified with different electroactive metals were synthesized: cadmium, bismuth, copper, silver  
135 and mercury. These metals are easily measured by anodic stripping voltammetry, and the stripping  
136 processes usually occur at different potentials. In order to study the electroactivity of these  
137 nanoparticles, cyclic voltammetry was employed with an electrolyte valid for the different metals:  
138 0.1 M pH 4.5 acetate buffer solution. The surface of the screen-printed working electrode was  
139 modified with an aqueous suspension of nanoparticles (2  $\mu$ L of 2 mg/mL). **Figures 1A** and **1B**  
140 show the cyclic voltammograms obtained for all the MTiPNPs. Although, the cathodic processes  
141 are difficult to assign to the reduction of metals since the oxygen reduction reaction takes place at  
142 close potentials and could affect in the response obtained, the most interesting process is the anodic  
143 stripping. Most of the nanoparticles showed one good resolved stripping peak due to the oxidation  
144 of the previously reduced metal. For BiTiPNPs, no stripping process was observed initially. To  
145 confirm the presence of bismuth into the nanoparticles, a preconcentration step was applied (-1.4 V  
146 for 30 s), and then the scan was performed towards positive potentials, observing the stripping  
147 process. This fact suggests that a lower amount of bismuth is introduced into the nanoparticles or  
148 the extraction is more difficult than for other metals (probably due to the lower solubility of  
149 bismuth cations). For CdTiPNPs, the cyclic voltammogram was recorded up to -1.4 V in order to  
150 achieve the reduction and observe the stripping process that appears at more negative potentials.  
151 These studies show that the kind of metal introduced into the MTiPNPs can be tuned and that has a  
152 strong influence in the detection of the nanoparticles.

153

154

155 It is expected that the cation exchange reaction leading to the metallic nanoparticles follows the  
156 general mechanism of the **equation 1**. This mechanism suggests that a higher extraction of protons  
157 from the initial nanoparticles would lead to a higher amount of metal introduced into the  
158 nanoparticle structure. Therefore, if a salt with a weaker base counter anion is used, it should be  
159 able to bind more protons, and to induce a shift of the reaction to the products according to the Le  
160 Chatelier's principle.



162 The effect of two different anions on the electrochemical response of CdTiPNPs was evaluated by  
163 using cadmium nitrate and acetate salts during the cation exchange reaction. After obtaining the  
164 final product as described in the Experiment section, 2  $\mu$ L of a 2 mg/mL aqueous dispersion of the  
165 nanoparticles were adsorbed into the working electrode, and square-wave anodic stripping  
166 voltammetry was used to carry out the detection of the metal from the nanoparticles (0.1 M pH 4.5  
167 acetate buffer was used). **Figure 1C** shows that the highest signal was obtained for the CdTiPNPs  
168 synthesized with acetate, and the lowest signal was found for CdTiPNPs synthesized with nitrate, as  
169 expected theoretically. These results show that the amount of metal can be tuned by using different  
170 anions in the cationic exchange reaction, and a higher amount of metals would lead to a more  
171 sensitive detection using electrochemical techniques. In previous works reported in the literature,  
172 only nitrate was employed for the cation exchange reaction, which suggests that the detection of  
173 these nanoparticles would be less sensitive than for those synthesized in this work.

174 [FIGURE 1]

175

### 176 **Enhancing the extraction of metals and detection of titanium phosphate nanoparticles**

177 As the introduction of metals is carried out by a cation-exchange reaction, the extraction of metals  
178 and their detection could be enhanced by reversing this reaction. In order to study this process,  
179 several electrolytic media were evaluated for the detection of the different MTiPNPs. 2  $\mu$ L of  
180 MTiPNPs aqueous solutions were added to the electrode surface and left to dry. Then, the cyclic

181 voltammograms were registered and the stripping peak currents were compared. **Figure 2A** shows  
182 the significant differences obtained for the same nanoparticles in different media or for different  
183 nanoparticles in the same medium. As a general rule, acidic media provided the best results for the  
184 metallic extraction and electrochemical detection, even obtaining a significant signal for BiTiPNPs  
185 in HCl and HNO<sub>3</sub>, which clearly demonstrates that acid media are able to extract more easily the  
186 metals from the nanoparticles. We have studied the cation exchange and the reverse process  
187 between TiPNPs and metals previously[23]. In this study, we showed that the metallic cations  
188 interact with the phosphate groups of the nanoparticles, and after acidic treatment, the metals are  
189 again extracted to the solution and the phosphate groups are recovered (as confirmed by the IR  
190 spectra). In previous works[14–17], acetate buffer was employed for the detection of the cadmium-  
191 based nanoparticles, and as we clearly demonstrate here, they are not the most appropriate metal or  
192 conditions to obtain a sensitive detection. Considering the peak currents (for sensitivity) and  
193 potentials (for selectivity in a multiplexing approach), H<sub>2</sub>SO<sub>4</sub> was chosen as the electrolytic  
194 medium for the following experiments.

195 [FIGURE 2]

196

### 197 **Multiplexing capabilities of metal-modified titanium phosphate nanoparticles**

198 The multiplexing properties of MTiPNPs were evaluated by square-wave voltammetry using 0.1 M  
199 H<sub>2</sub>SO<sub>4</sub> as the electrolytic medium. Electrode surface was modified with 2 μL of 0.5 mg/mL  
200 MTiPNPs of different binary mixtures (and individually to assign each stripping peak). **Figure 2B**  
201 shows the voltammograms of individual MTiPNPs and **Figure 2C** shows the voltammograms of  
202 several mixtures. Peak potentials of the stripping processes were in the following order: BiTiPNPs  
203 (-0.35 V), CuTiPNPs (-0.25 V), AgTiPNPs (-0.1 V) and HgTiPNPs (+0.2 V), while that a very  
204 small peak was observed for individual CdTiPNPs around -1 V (it increased in some of the binary  
205 mixtures (in presence of Hg and Bi)). Under these conditions, the stripping of BiTiPNPs/CuTiPNPs  
206 and AgTiPNPs/HgTiPNPs could not be resolved. Considering these facts and the magnitude of the

207 peak currents, CuTiPNPs and HgTiPNPs were chosen for the multiplexing experiments because the  
208 stripping processes were perfectly resolved and a high peak current was obtained for both  
209 nanoparticles.

210

211 In order to evaluate the analytical performance of the Cu and Hg-modified titanium phosphate  
212 nanoparticles, voltammograms were recorded after the modification of the electrode surface with  
213 increasing concentrations of nanoparticle mixtures (2  $\mu\text{L}$  solution). A potential of -1 V for 30 s was  
214 chosen as the optimal electrodeposition step. **Figure 3A** shows the voltammograms obtained for  
215 increasing concentrations of HgTiPNPs and CuTiPNPs. A working linear range between 0.01 and  
216 0.3  $\mu\text{g/mL}$  was obtained for CuTiPNPs, whereas a linear range between 0.01 and 0.4  $\mu\text{g/mL}$  for  
217 HgTiPNPs was obtained. The response was linear according to the following equations:  $i_p$  ( $\mu\text{A}$ ) =  
218  $133 (\pm 5) [\text{CuTiPNPs}] (\mu\text{g/mL}) - 0.1 (\pm 0.2)$ , ( $R^2 = 0.992$ ) and  $i_p$  ( $\mu\text{A}$ ) =  $193 (\pm 1) [\text{HgTiPNPs}]$   
219  $(\mu\text{g/mL}) + 0.5 (\pm 0.1)$ , ( $R^2 = 0.9994$ ), with estimated detection limits of 0.01 and 0.008  $\mu\text{g/mL}$ ,  
220 respectively. Clearly, the greater amount of metal introduced into these nanoparticles and the most  
221 effective detection allowed their determination at much lower concentrations than for previously  
222 evaluated AgTiPNPs[19] or CdTiPNPs[21], even with lower deposition times, leading to  
223 nanoparticles with very promising multiplexing properties. These results suggest that HgTiPNPs  
224 and CuTiPNPs could enhance significantly the detection in biosensing when these nanoparticles are  
225 used as labels in comparison to the typically used CdTiPNPs[14–17].

226

227 Copper and mercury-modified titanium phosphate nanoparticles were modified with neutravidin  
228 following a method previously reported in the literature[23], and applied, as a proof-of-concept, in a  
229 multiplexing biosensor for the simultaneous detection of anti-transglutaminase IgG and IgA  
230 antibodies. Most of the biosensing steps were similar to those described in a previous work[22], but  
231 CuTiPNPs and HgTiPNPs were bound to anti-IgG-BT and anti-IgA-BT, respectively, by the  
232 neutravidin-biotin interaction. A mixture of the positive and negative serum controls of two

233 commercial ELISA kits for anti-tTG IgA and IgG detection was used as sample solution (1:1:2 ratio  
234 for controls and PBS). **Figure 3B** shows the voltammetric response obtained for 0, 7.3 and 45.5  
235 U/mL concentrations of the analyte mixture, which demonstrates the good analytical performance  
236 of the multiplexing approach using CuTiPNPs and HgTiPNPs.

237 [FIGURE 3]

238

## 239 CONCLUSIONS

240 In this work, we have shown the possibility of tuning the amount of metal introduced into titanium  
241 phosphate nanoparticles by varying the type of metal and the counter-ion used during the cation  
242 exchange reaction. This fact has allowed to obtain nanoparticles with a great metallic load, and,  
243 therefore, with an excellent detection capacity for their use as detection label in electrochemical  
244 biosensors. In addition, it has been demonstrated that the electrolytic medium used in the detection  
245 step plays a crucial role for the metal extraction and its correct choice has allowed the very sensitive  
246 detection of the nanoparticles. These results have provided the basis for the development of a  
247 detection method for these metal-based nanoparticles, which were used successfully as detection  
248 label in a multiplexing electrochemical biosensor.

249

## 250 ACKNOWLEDGEMENTS

251 This work has been supported by the FC-15-GRUPIN-021 project from the Asturias Regional  
252 Government and the CTQ2014-58826-R project from the Spanish Ministry of Economy and  
253 Competitiveness (MEC).

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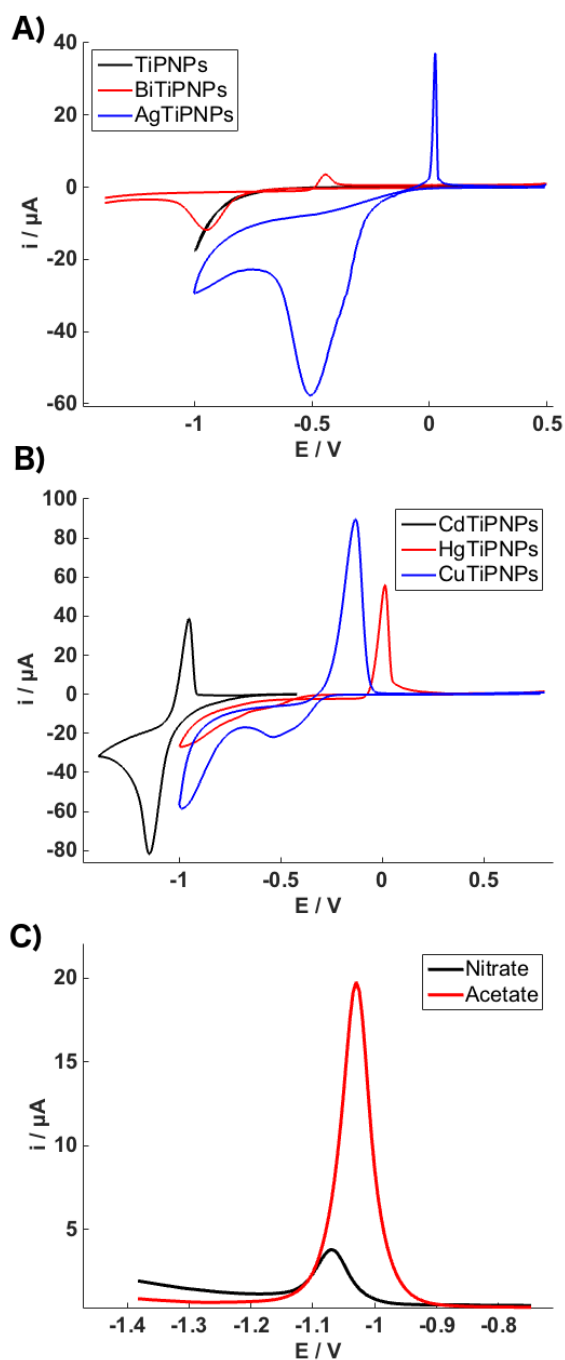
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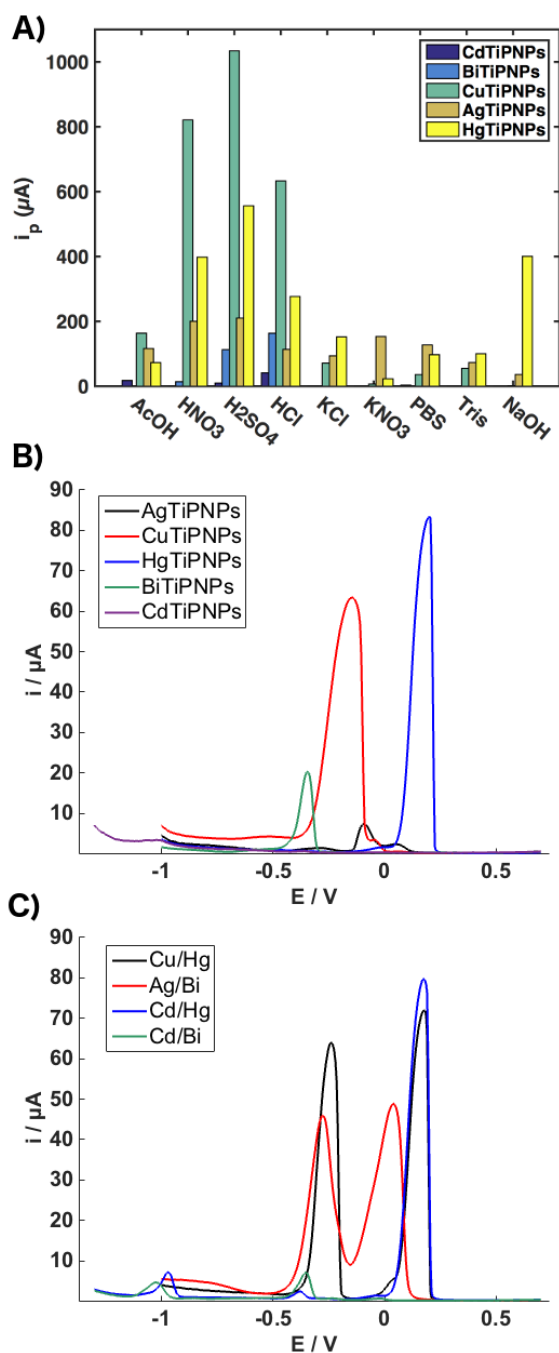


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328 **Figure 1.** A) Cyclic voltammograms of TiPNPs, AgTiPNPs (between +0.5 and -1.0 V) and  
329 BiTiPNPs (between -1.4 V to +0.5 V with a deposition step at -1.4 V for 30 s). B) Cyclic  
330 voltammograms of CdTiPNPs (between -0.4 and -1.4 V), HgTiPNPs and CuTiPNPs (between +0.8  
331 V and -1.0 V) in 0.1 M pH 4.5 acetate buffer. C) Square-wave voltammetry of CdTiPNPs  
332 synthesized by using a nitrate or acetate salt during the cation-exchange reaction.

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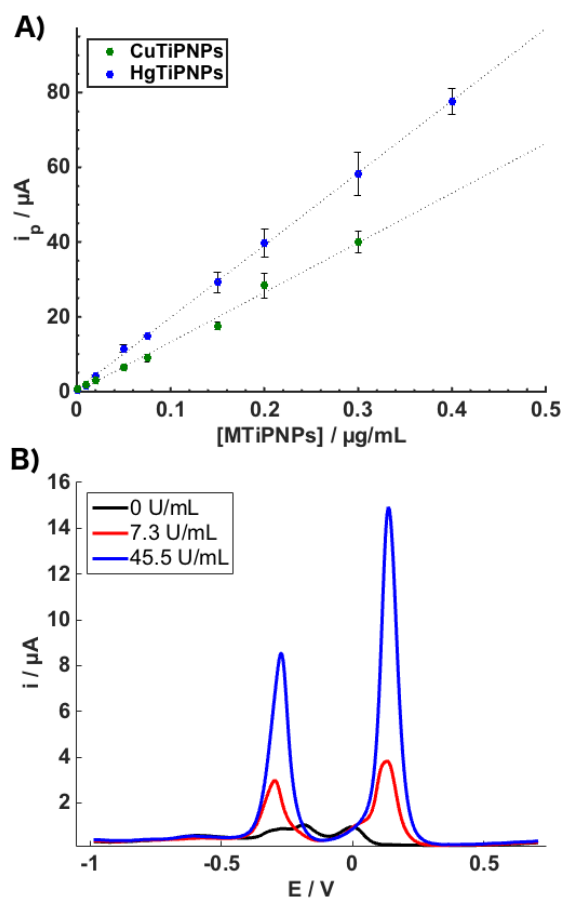




335

336 **Figure 2.** A) Peak currents of the anodic stripping process for the different MTiPNPs using  
 337 different electrolytic media. Concentration of electrolytes was 0.1 M. B) Square-wave voltammetry  
 338 of the different MTiPNPs in 0.1 M H<sub>2</sub>SO<sub>4</sub>. C) Square-wave voltammetry of several MTiPNPs  
 339 binary mixtures in 0.1 M H<sub>2</sub>SO<sub>4</sub>.

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342 **Figure 3.** **A)** Calibration plots of the HgTiPNPs and CuTiPNPs. **B)** Electrochemical response of the  
 343 proof-of-concept multiplexing biosensor using HgTiPNPs and CuTiPNPs as detection label.

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